A ring-closing metathesis approach to a synthesis of the B ring of eleutherobin

Krishna P. Kaliappan* and Nirmal Kumar

Department of Chemistry, Indian Institute of Technology-Bombay, Powai, 400 076 Mumbai, India

Abstract—A short and efficient RCM route is reported for the construction of the key nine-membered B ring of eleutherobin starting from the readily available 1,2,5,6-diisopropylidene-D-glucose.

The success of taxol[®] (paclitaxel) against breast and ovarian cancer has elicited a worldwide search for better sources and improved analogues of this drug which might operate through similar modes of action.¹ This led to the discovery of the new natural products discodermolide, epothilones and eleutherobin which displaying a paclitaxel-like mechanism of action. Among them, the latest discovery, eleutherobin **1**² was isolated from a marine soft-coral found in the Indian Ocean and found to be closely related to sarcodictyins **2**, **3**³ and valdivone **4**.⁴ Eleutherobin has been shown to possess potent cytotoxicity with an IC₅₀ of 10.7 nM which is comparable to taxol. As eleutherobin was only available in scarce amounts from natural sources, chemical syntheses have become more imperative to study its biological properties further. This prompted synthetic chemists to develop new strategies⁵ to synthesize eleutherobin and their sustained efforts have culminated in two total syntheses.^{6,7} However, convenient access to the fully functionalized nine-membered B ring remains a challenge in the synthesis of eleutherobin.

As a part of our program directed towards the development of simple and efficient routes to the syntheses of tubulin binding anticancer agents, we became interested in the synthesis of eleutherobin and its congeners. A strategy for the synthesis of eleutherobin was designed based on the retrosynthetic analysis shown in Scheme 1. We envisaged that a ring-closing metathesis (RCM) reaction⁸ would be an ideal key reaction to construct these natural product skeletons. Eleutherobin possesses

three double bonds, however to be formed as a result of an RCM reaction, only disconnection of the B ring double bond will simplify the target molecule leading to precursor **C** which then can be further disconnected to the bicyclic enone **7** and the epoxide **8**. It is clear that the success of our synthetic route to eleutherobin depends heavily on the construction of the B ring with the strategically placed double bond. So, a model study was carried out to synthesize the highly oxygenated nine-membered ether ring⁹ starting from the readily available 1,2,5,6-diisopropylidene-D-glucose and herein we report our initial results.

Our synthesis, as depicted in Scheme 2, starts with the classical etherification of the secondary alcohol **9** to afford **10**. Under mild acidic conditions, the more exposed 5,6-*O*-isopropylidene group was selectively

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Scheme 1.

Scheme 2. Reagents and conditions: (a) NaH, allylbromide, THF, TBAI (cat.), rt, 2 h, 93%; (b) 60% aq. AcOH, H₂O, 12 h, 90%; (c) silica gel supp. NaIO₄, CH₂Cl₂, 30 min, 86%; (d) 4-bromo-1-butene, Mg, THF, 0°C to rt, 12 h, 83%; (e) NaH, MeI, 0°C to rt, 2 h, 77%; (f) TBSCl, imid., DMF, 0°C to rt, 10 h, 84%; (g) **18** (10 mol%), CH₂Cl₂, 40°C, 2 h, (33% for R = Me, 30% for R = TBS); (h) **18** (10 mol%), Ti(OiPr)₄ (7.5 mol%), CH₂Cl₂, 40°C (63% for R = Me, 83% for R = TBS).

deprotected to give a diol which was subsequently cleaved by treating with silica gel supported NaIO₄¹⁰ to afford **11**. Addition of the Grignard reagent, derived from 4-bromo-1-butene and magnesium, to the aldehyde **11** afforded two diastereomeric products **12** and **13** as a readily separable mixture in 3:2 ratio¹¹ and good chemical yield. Subsequent protection of the major alcohol **12** as its methyl ether set the stage for the key RCM reaction. Unfortunately, when the RCM was carried out with Grubbs catalyst **18** under high dilution conditions (0.003 M solution with 10% catalyst), only 33% of the desired cyclized product **16** was obtained, the remainder being unreacted starting material and presumably some cross-metathesis product. The observation of a similar result with the TBS ether **15**, prepared from **12**, suggested that the low yield could be attributed to coordination of the metal centre with the oxygen of the furanose ring.¹² In

order to destabilize this chelate structure, the RCM reaction of the dienes **14** and **15** was carried out with a catalytic amount of **18** in the presence of a sub-stoichiometric amount of Ti(OiPr)₄¹³ and this modified protocol successfully led to the formation of the desired RCM products **16** and **17**, respectively in high yields.¹⁴

In summary, a seven-step synthesis of a highly oxygenated model of the key nine-membered B ring of eleutherobin was successfully achieved from commercially available D-glucose. Interestingly, the RCM products **16** and **17** possess all the structural features of the marine natural products isolaurallene **4**¹⁵ and neolaurallene **5**.¹⁶ We are currently examining the extension of this methodology, in our laboratory, to synthesize these natural products, in addition to eleutherobin and its congeners.

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- All compounds reported here were duly characterized. Selected data: Compound **16**: $R_f=0.4$ [silica gel, ethyl acetate:hexane (4:1)], $[\alpha]_D -63.9$ (c 1.0 CHCl_3); IR: (neat, cm^{-1}): 2940, 1657, 1466; ^1H NMR (300 MHz, CDCl_3): δ 5.89 (d, $J=3.8$ Hz, 1H), 5.83 (q, $J=9.2$ Hz, 1H), 5.48 (dt, $J=9.4$, 4.2 Hz, 1H), 4.43 (d, $J=3.8$ Hz, 1H), 4.31 (dd, $J=14.1$, 3.8 Hz, 1H), 4.06 (d, $J=3.1$ Hz, 1H), 4.01–3.51 (m, 2H), 3.37 (s, 3H), 3.37–3.32 (m, 1H), 2.34–2.13 (m, 2H), 1.94–1.83 (m, 1H), 1.41 (s, 3H), 1.40–1.31 (m, 1H), 1.25 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 135.6, 125.6, 111.5, 104.8, 85.3, 83.4, 82.5, 80.5, 65.9, 57.5, 27.9, 26.8, 26.4, 26.0. Compound **17**: $R_f=0.8$ [silica gel, ethyl acetate:hexane (4:1)], $[\alpha]_D -39.9$ (c 1.0 CHCl_3); IR: (neat, cm^{-1}): 2947, 1650, 1255, 1091; ^1H NMR (300 MHz, CDCl_3): δ 5.9 (d, $J=3.7$ Hz, 1H), 5.84 (q, $J=9.1$ Hz, 1H), 5.52 (dt, $J=10.2$, 5.1 Hz, 1H), 4.47 (d, $J=3.7$ Hz, 1H), 4.35 (dd, $J=14.2$, 3.7 Hz, 1H), 4.08 (d, $J=2.9$ Hz, 1H), 3.96–3.82 (m, 3H), 2.39–2.14 (m, 2H), 1.91–1.51 (m, 2H), 1.47 (s, 3H), 1.30 (s, 3H), 0.88 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 135.9, 125.7, 111.3, 104.6, 86.4, 84.1, 81.1, 73.8, 66.2, 33.2, 29.8, 26.9, 26.5, 26.0, 18.4, –4.3, –4.8.
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